

DNA Based Molecular Detection Methods of Biological Adulterations in Commercial Teas: Successes and Obstacles



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Abstract: Extra to water, tea is the broadly consumable hot beverage in the biosphere. People drink more tea than coffee, cold drinks, dairy products, and alcohol combined. Availability does not effectively order the dried plant pieces, which are utilized to make tea. Numerous cases of food adulteration have been recorded in many countries, including India. This has taken to the creation of a new discipline of science, known as ‘green criminology’, to compensate violations of food law. Over the years, new techniques for identifying food adulteration have been developed Here we reviewed different standard DNA based molecular techniques to analyze their performance in identifying tea constituents. Originally, these were sensory techniques, which proved unreliable. Later, physical analysis of the product was done based on data on the label and microscopic evaluation. Later methods, based on the identification of lipids and proteins, were also not accurate due to biochemical changes during processing. These problems provoked scientists to get an interest in the potential of DNA testing. Due to the stability of DNA and the universal applicability of DNA-based methods for all cells, they are ideal for use in practice. Currently, the most reliable test for detecting food adulteration is Bar-HRM, as it is a highly sensitive and specific technique. Broad scale endorsement of the plant’s DNA barcoding may require systems that spot the journey brings about point of view of common plant species names and character for separate firmly close gatherings.

Keywords: Biological Adulterations, Camellia Sinensis, Tea, DNA Barcoding, HRM analysis, DNA based Molecular Modeling and Detection Techniques, Food Adulteration.

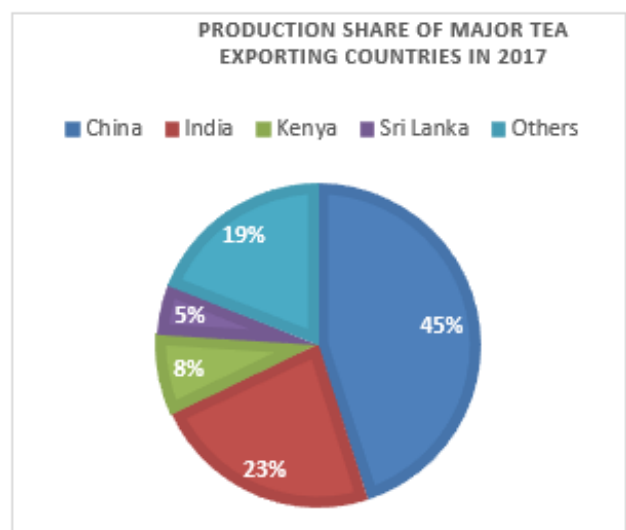
I. INTRODUCTION:

“Chai can’t be described by one word. Spicy, Malty, Earthy- the list is never-ending. Add a dust of cardamom, a hind of ginger, sprinkle some cinnamon, the possibilities are endless.

And the important ingredient that makes this brew special and versatile is Indian Black Tea. So, the next time you’re in any mood for that matter, grab a cup of Indian masala chai”. - is described in the Tea board of India. Overall the landmass aside from North America with the expressive scope of agro-climatic conditions among Georgia and Argentina, more than 36 countries spread grow tea. In the year 2017, the earth tea production and consumption were 812 million Kg and around 5177 million Kg respectively. During 2017, the total export from producing countries added up to 1796 million Kg. China, India, Kenya, and Sri Lanka are the primary tea harvesting importing and exporting nations on the planet and they represent 76% and 73% of the world harvesting and exporting respectively[1].

Table 1. Production and export share of the Major Tea Producing And Exporting Countries During 2017 [1].

Country	Harvesting		Export	
	Million (Kg)	Global share (%)	Million (Kg)	Global share (%)
China	0.2609	0.45	0.355	0.2
India	1322	0.23	0.252	0.14
Kenya	0.44	0.8	0.416	0.23
Sri Lanka	0.308	0.5	0.286	0.16
Others	0.1133	0.19	0.487	0.27
World Total	0.5812	0.1	0.1796	0.1



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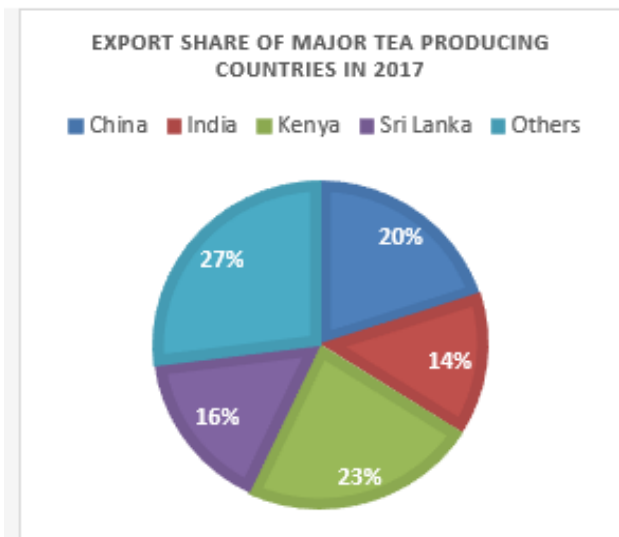
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Graph 1. Graphical representation of Production and export share of the major tea producing and exporting countries during 2017 [1].

Out of 15 tea cultivating states of India: Assam, West Bengal, Tamil Nadu, and Kerala are the main tea cultivating states. These states contribute about 98% of the total tea harvesting. The world's finest tea like Darjeeling, Nilgiris, Assam, and Kangra, which are legendary for their gentle flavor, strength, and intensity, are produced in India. Year 2017-18, is a record year of tea production with an increase of 74.65 M. Kg over 2016-17 due to enhanced climate conditions that are prevailing in the principle tea cultivating zones both in North and in South India. India registered the highest ever tea exports during the year 2017-18. It was higher by 29.46 M Kg and 28.94 M Kg in volume terms and 519.48 Crs and 432.38 Crs in volume terms during the financial years over the corresponding period last year. The anticipated national retention of tea for the year 2017-18 was round 1089 M.Kg as against 1044 M.Kgs in 2016-17 [1] Watery imburements arranged from dried plants, comprehensively known as tea, famous drink with attractive physiological activities, and potential medical advantages. Tea appropriately alludes to imburements arranged from parts of the *Camellia sinensis*, a perennial blooming tree in the family Theaceae, local to the dry areas of south-western China and adjacent nations [17, 53]. The two primary business assortments are little leafed *C. sinensis* var. *sinensis*, adjusted to warm climate and high height, and enormous leafed *C. sinensis* var. *assamica* (J. W. Pole.) Kitam., which develops healthily in sub-tropical, tropical, and mangrove conditions.

Tea leaves consist of a good proposition of phytochemicals, comprising methylxanthine, caffeine and the polyphenolic catechins [5, 37, 83]. Drinking tea as a hot beverage began in southern China at any rate 2000 years prior, and today tea is the broadly expended refreshment on the planet [16,44]. Various methodologies, from drying and annealing to long periods of microbial maturation, produce the assortment of tea types; white, green, dark, oolong, and pu-erh, which contrast in catechin substance and antioxidant action. [2,44,48] Exact naming is significant for customers, advertisers, and controllers as tea composition can't be effectively distinguished to species by physical appearance [78]. Their ordered assorted variety and aromatic nature present a prepared and requesting trial of DNA-based

distinguishing proof [28]. Here we review the success and obstacles to distinguishing tea fixings utilizing various DNA based procedures including, short DNA sequencing from the uniform region inside the genome, DNA barcoding, high-resolution melting curve analysis, and molecular modeling. Food corruption is the purposeful or accidental incorporation of substances that are not lawfully endorsed in improvement, prompting an impersonation of the item and a very high decrease in market esteem [2, 49]. Financially accessible plant items are defenseless to replacement or pollution with other materials. This leads to a decrease in the excellence of the product and may provoke inebriation and hypersensitivity. Deliberate or monetary defilement is happening to emulate enhanced organoleptic and visual quality by the expansion of outside materials. On the other hand, for example, botanical fraudulent, waste segments including counterfeit colorants, spent concentrates, and plant portions of various unrelated species are the regularly distinguished adulterants [3, 68]. FSSAI defined "adulterant" as any component which is or could be utilized for making the food hazardous, inadequate, mis-marked, consisting of the superfluous issue. As per FDCA, the essential sanitation law should be controlled by the FDA. Food can be announced adulterated if there is:

- Blending of an unrelated substance, which is damaging to health.
- Blending of the cheap or low-grade quality item to food.
- Any esteemed constituent is expelled from fundamental food source
- Less standard quality food.
- Any constituent rises the mass or weight.
- Material that makes it appear more valuable

So, as to guarantee food safety and quality, different strategies are utilized to recognize and evaluate adulterant in tea. A few instrumental strategies have been utilized and with progressions in diagnostic science, most current techniques are known for their factors, exactness, selectivity, and quickness.

As of late, high delicate DNA based sub-atomic methods like HRM investigation are additionally being investigated for recognition of adulterants in tea products.

II. SOURCES OF TEA POLLUTION

A. Metallic contamination

The metals and metal compounds are accidentally included along with the food products which are maybe from the seashore or agricultural example tea, oil foods, and herbal medicinal plants. Mostly Cadmium, Lead, and Copper are the most abundantly present in the food source [47].

B. Microbial contamination

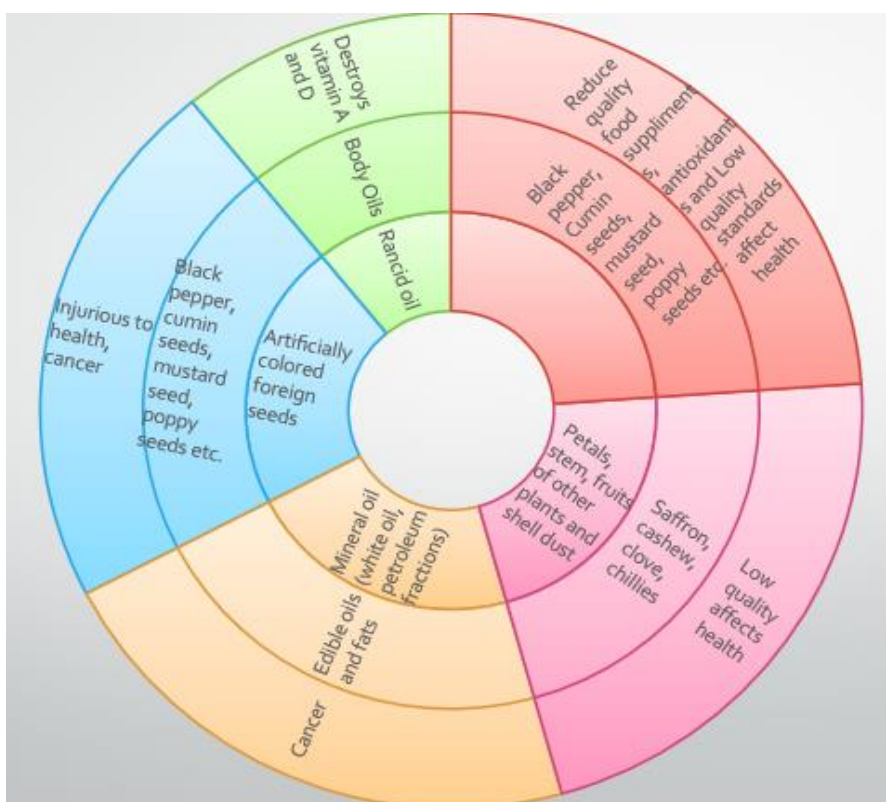
It is defined as the contamination of tea because of the blend of different microorganisms through a different premise. Microorganisms whenever from numerous sources might corrupt nourishment during tea processing like during Picking, Withering, Rolling/Brusing, Fermentation, Fixation, Drying, and Packing and Store [20].



C. Health risks of adulterated tea

Tea defilement contributes to the general public with numerous infections running from mild to harsh extremes like vision issues, liver issues, skin sicknesses, and a few stomach issues, for example, the runs [8] Ordinarily, observed health issues are asthma, skin infections and malignant growth caused because of ingest fish, natural products, meat defiled with synthetic substances like formalin. Health is exceptionally delicate to adulterants and once in a while shows prompt symptoms like looseness of the bowels, diarrhea, and vomiting. For instance, turmeric powder subbed wit tamarind or date seed powder can cause looseness of the bowels [27]. Debasement of cream filed nourishment, bread shop things, and dairy items can

likewise cause stomach issues and regurgitating [4]. Inappropriately handled canned meat and milk may cause food contamination and stomach torment or other food diseases as a rule with fever and chills. Notwithstanding quick impacts, there might be some drawn-out unfavorable impacts of debased food. Long impacts like colon, peptic ulcers, liver infections like cirrhosis and liver disappointment, heart sicknesses, a blood issue, bone marrow variation from the norm, and kidney harm have been seen because of adulterants like shading colors, calcium carbide, urea, consumed motor oil and some of the time even because of abundance measure of allowed additives [52]. Some of the biological adulteration and their health impacts are given in Graph 2.



Graph 2. Health effects of common biological adulteration in food [72].

III. MOLECULAR METHODS FOR DETECTION OF TEA ADULTRATION:

A. Random amplified polymorphic DNA (RAPD);

The primers are evolving by length stand generally a single primer, which ties to inverse strands of DNA, is utilized[57]. Since the base gets fixed to different locales on the DNA particle, various sections of DNA are enhanced. The output is variable-sized groups, which are perceptible on EtBr. staining the gel. The patterns of DNA groups are normally distinctive for a plant species.

At the point when distinctive plant species are compared, a few bands will either be available or missing, in this way creating a polymorphism. Habitually, numerous RAPD primers are utilized to identify polymorphisms, particularly between related plants. RAPD markers in this manner give an innovatively basic and generally speedy approach to identify genetic fluctuation, which can frequently be seen at the basic level.

For instance, RAPDs and related techniques have been utilized for distinguishing genetic variation in various promotions of green tea and sweet potato [67, 88]. Nevertheless, a considerable lot of these methods have gotten less across the board for a multi-decade. There are a few purposes behind this.

- There is inadequate amplification of DNAs fetched about by antagonism for primer sites, because of low annealing temperature [91].
- The shortage of productivity from lab to lab, due to changes in PCR considerations [85].
- It is not conceivable to classify a completely unfamiliar and unexpected adulterant.

B. Amplified Fragment Length Polymorphism.

AFLP collaborates with RFLP, and was created as overarching methods for DNA fingerprinting [87]. In population studies AFLP is an appreciated tool because it can perceive genomic variation even in the tiny of requiring sequence data, the AFLP can perceive genomic variations; hence, it has been appreciated as a tool in the population studies. Through the sequence of bands, various models can be scrutinized for polymorphisms, they can be from the variant in restriction sites, the occurrence of deletions or insertions, and/or nucleotide alterations next to the restriction site[60] AFLP method is more convenient for basic methods rather than species discrimination, more particularly if the species are closely linked. High quality DNA is mandatory, a second drawback is difficult to obtain from processed, dried teas [99]. Moreover, AFLP is required sound technical expertise for reliable results and costly [24, 37, 79, 99].

C. Specific-Specific DNA Markers

An inconvenience to AFLP and RAPD analysis the capacity to recognize variety within a species makes it hard to create outlines that will be related to the species overall [32]. In this way, keeping DNA analysis to a strategy known locus of enthusiasm rather than irregular obscure loci may have more prominent utilization when species recognizable proof is wanted [76]. To effectively describe species dependent on

DNA standardized tags, the determination of useful DNA districts is significant. A decent DNA marker ought to have low intra-explicit and high between explicit changeability [28]. Different types of molecular marker measures have been demonstrated to help recognize plants at the class and even at the species level. These DNA locales naturally have fragments that are saved enough for general primers to be planned, yet have areas that advance moderately quickly with the goal that the intensified district is species-level variation studies. One model is the ITS (Internal Transcribed Spacer) locale contained inside the 18S -26S, ribosomal DNA (rDNA). The ITS can be distinct all-around any plant species since amplification can be intended for the very much moderated 18S and 26S qualities [6, 9]. General ground works have additionally been depicted for the focal 5.8S rRNA quality, and regularly the shorter ITS-1 or ITS-2 areas are utilized for examination. Different groupings in ribosomal RNA DNA have additionally been utilized to confirm herbals, including the 18S RNA DNA arrangement [12] and the chloroplast 5S area and spacers [45, 84]. Chloroplast qualities, in particular rbcL and trnK, just as matK, a quality contained inside trnK, all of which have been utilized in phylogenetic examinations of plants, are likewise used to deliver atomic fingerprints of restorative plants [12]. matK is a quickly developing quality in the chloroplast genome [35], and may not show critical contrasts between firmly related species.

b. Species-specific PCR

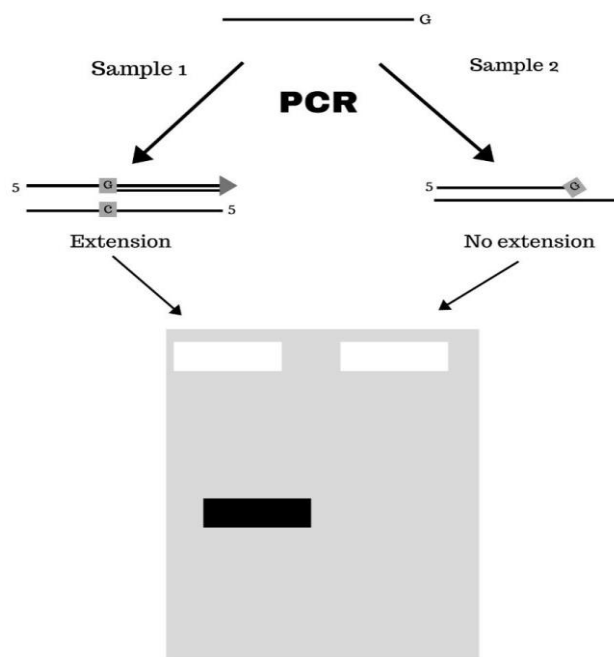


Figure 1:Species-specific PCR[Created by authors]:

A primer structured that is totally reciprocal to sample 1, however has a 3'- crisscross to non-target species, for example, sample 2. At the point when utilized in PCR, just DNA from tests that strengthen and have ideal matches to the 3'- end of the primer will be intensified. In this way, a PCR item for sample 1, however not sample 2, will be seen on an agarose gel.

D. Direct Sequencing of PCR Products

Routine DNA-based strategies, for example, RAPD, AFLP, AP-PCR, Specific DNA Markers, and SSR/SNPs experience the ill effects of the absence of consistency and lacking normalization [22].



These issues have provoked the improvement of extensively available instruments, for example, DNA barcoding to distinguish item replacement and to confirm commercial items[62]. Amplification of the ITS area with the help of conventional PCRA technique, separating the bands through gel separation process of each sample, followed by straight sequencing of the isolated sample and parallel correlation with the NCBI database will be performed to affirm the presence of plant-based adulteration [41, 54] in commercial teas.

Be that as it may, this sort of analysis might be ruined by the absence of defiling DNA. This can create a PCR product that is hard to investigate [77]. The presence of polluting plant species may likewise confound such an examination. For example, when we analyzed contamination in the marketed tea powder, which had been isolated from packed tea powder, we observed that sequencing of the ITS PCR Product without cloning generated mix picks. [48] propose a potential clarification was that there was a more prominent match between the all universal primer destinations and the ITS locale different types of camellia plants. Then again, contrasts in ITS copy number and genome size may have brought about a special amplification of different plants over *Camellia sinensis* DNA in the tea powder.

E. Cloning of PCR Products Detects Contaminants

On the off chance that there are different species present along with the known plants, these would not be recognized except if a portion of the RAPD-determined fragment is exposed to advance analysis, for example, sequencing. And still, at the end of the day, this would be a troublesome

endeavor since discovering contaminant DNA amidst all the bands will be like searching for a "needle in a haystack". As talked about above, direct sequencing of specific marker sequences additionally doesn't uncover the presence of fraud species. Before going to sequencing performing cloning the PCR product and then sequencing the clones are one approach to test the composition of a PCR product that might be derived from various species [48]. The number of clones is representative of the diversity found in the sample is a big troubleshoot. Besides, cloning followed by PCR is time-consuming and expensive.

IV. PCR-RFLP

RFLP is the aftereffect of a restriction enzyme breakdown of DNA. These enzymes cut DNA at explicit sites comprising of a few nucleotide; a solitary nucleotide distinction can bring about the enzyme not perceiving the site in the DNA and subsequently not cutting it (Fig. 1a). If two plant species vary in the site where a specific enzyme confines the DNA, the outcome is an example of DNA bands of various sizes. In this novel method, the PCR fragment is the breakdown by a specific restriction enzyme yielding at least two sections, which are identified on an EtBr. stained agarose gel. If adulteration is present, and the enzyme does not hamper its DNA, then the PCR fragment originated from the impurity DNA will not be cut. Fig 1a. [21, 48, 100]. If in case of the DNA sequence is not known, then the PCR-RFLP technique is failed to analyze. The enzymes that should be used must be sound in quality; otherwise, the adulterant DNA reveals the same restriction site as the DNA isolated from the teas.

a. RFLP

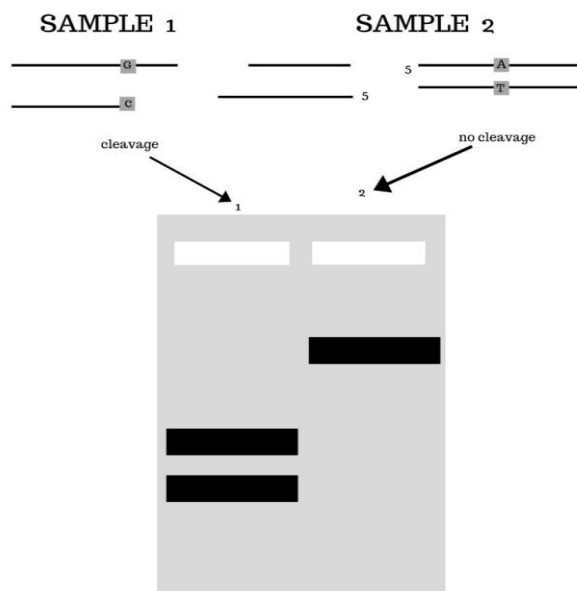


Figure 2: Restriction fragment length polymorphism (RFLP) [Created by authors]: A particular restriction endonuclease is picked that cuts a DNA marker succession of sample 1. Nonetheless, a similar DNA marker from sample 2 isn't severed because of the nearness of a solitary nucleotide polymorphism that adjusts the acknowledgment site. After assimilation, an agarose gel will show the nearness of two DNA pieces for sample 1, however just a single higher atomic weight DNA section will be seen for sample 2.

A. Single-Strand Conformation Polymorphism

SSCP is going after the standard under non-denaturing conditions (single-deserted) ssDNA will get different compliance depending upon its nucleotide grouping (Fig. 1c). This system was created for the acknowledgment of SNPs in individuals and has been utilized for the distinguishing proof of genetic varieties related to affliction [26]. SSCP incorporates the PCR incorporates, the lesser the area, the improved the objectives [79]. Single-surrendered DNA is made from two fold scalation of a region of interest, regularly under 400 nucleotides inserted PCR either by lambda ex-nuclease or

by heat denaturation [64] to unequivocally degenerate the strand that has had a 5' phosphate gaveto it employing the primers [75].

The ssDNA is run on a non-denaturing gel and its change relocation can be imagined by recoloring with silver, EtBr. or fluorescent colors [85]. Furthermore, a corroborative advance should be possible by disengaging the DNA groups of enthusiasm from the gel, and afterward, re-enhancing followed by sequencing, lastly contrasting the DNA arrangement with the NCBI database or to a known succession [85]. This implies potential adulterants that don't coordinate the normal unique mark can be distinguished.

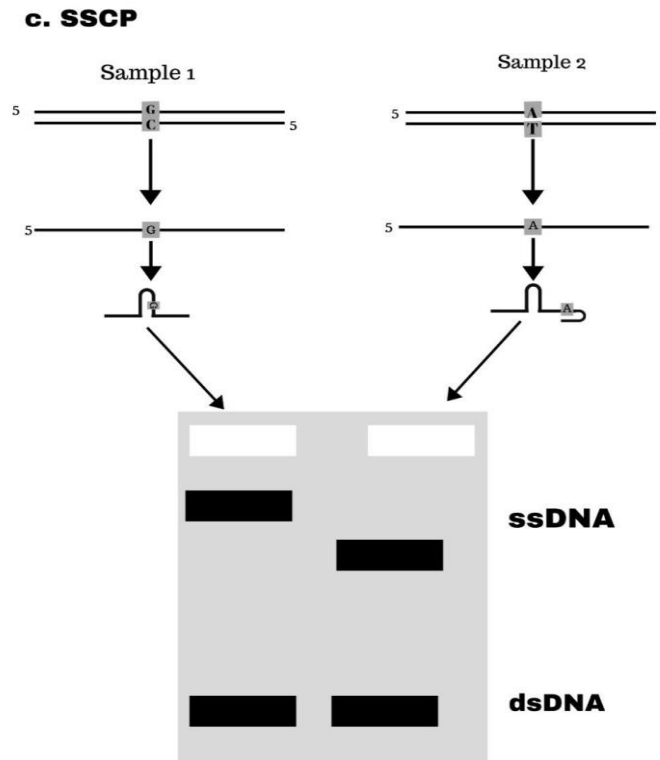


Figure 3: Single-strand conformation polymorphism (SSCP) [Created by authors]: A DNA marker sequence is enhanced by PCR and single-stranded (ss) DNA is then produced. ssDNA of sample 1 expect an auxiliary adaptation that varies from sample 2 because of variety in the succession. A distinction in relocation between the two samples will be identified on a non-denaturing polyacrylamide gel due to the diverse electrophoric mobilities of the two adaptations.

B. Microarrays

Microarrays offer a mechanical assembly for the screening of allelic assortment, it is a high throughput method for a review, see [10]. A hard substrate, generally silicon, is spotted with explicit single abandoned oligonucleotides that go an area covering the assortment. The chip is hybridized with stamped DNA. Hybridization is recognized with the help of fluorimetrically or colorimetrically. Without a doubt, differential hybridization can be achieved even if a single nucleotide change occurs in the sample. Microarray is a significant technique for the area of polymorphism studies (Fig. 1d) [25]. A couple of examiners have demonstrated that the development of microarray can be applied to the approval of plant species. In the year 2005 Carles et al., [14] have been successfully demonstrated DNA isolated

from twenty diverse plant species employing the 5S chloroplast ribosomal RNA quality as a templet. Every one of the twenty animal categories could be isolated, including people from comparable classes. A hindrance is that the objective arrangement ought to be known; in this way simply those DNA groupings speak to the chip will be recognized. Species that may be contained in the regular, in any case, are not spoken to the chip, will remain unobserved. It will be fundamental to recognize the allelic assortment that may occur inside the masses to guarantee that every variety of alleles spoken to. Microarrays, anyway suitable for screening solitary models for different allelic polymorphisms, are not as ideal for screening colossal amounts of tests on the chip that each chip can simply separate every model.

d. MICROARRAY

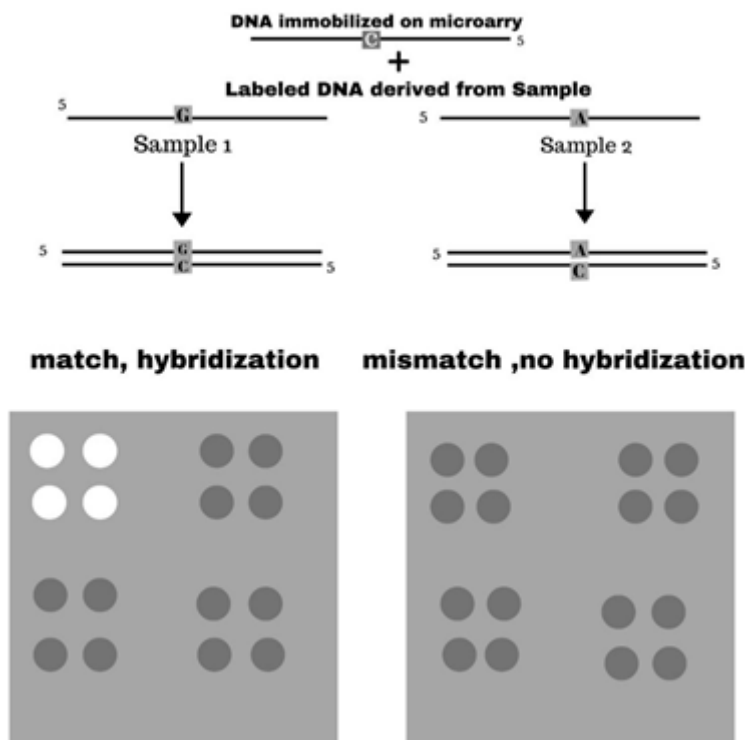


Figure 4: Microarray [Created by authors]: A DNA marker sequence from sample 1 is immobilized on a strong substrate. In spite of the fact that variations of this DNA marker are additionally present on the array, the sequence relating to sample 2 isn't. The DNA marker is intensified from DNA of sample 1 or sample 2 and named for use as a test for the array. States of brooding are balanced with the goal that solitary DNA that base combines impeccably is steady and can be identified. DNA from sample 1 is identified on spots relating to its species, while sample 2 isn't recognized in light of the fact that it isn't belongs to the array.

C. Denaturing Gradient Gel Electrophoresis Analysis of PCR (DGGE-PCR)

Separation of bands in DGGE depends on the electrophoretic portability of a partially melted DNA molecule in polyacrylamide gels, which is diminished contrasted and that of the helical type of the molecule. Stretches of base sets with a similar melting temperature. Succession variety inside such spaces makes their melting temperatures vary. sequence variations of specific sections will in this manner quite moving at various positions in the denaturing slope and consequently can be isolated successfully by DGGE fig. [40]. This procedure has been effectively applied to distinguishing sequence variations in various qualities from several unique living beings. DGGE can be utilized for direct investigation of genomic DNA from living beings with millions of base pairs[52].

This is finished by moving the partition to hybridization layers by capillary blotching with changed gel media [88] or by electroblotting [23] trailed by an investigation with DNA tests. On the other hand, PCR [71] can be utilized to specifically enhance the grouping of enthusiasm before DGGE is utilized [13].

In an alteration of the last strategy, GC-rich arrangements can be consolidated into one of the primers to change the melting nature of the fragment of interest, to the extent to which near 100% of all conceivable sequence varieties can be recognized [60, 78].

D. DNA Variety Test (DVT):

The technique of DNA Verity Test is appropriate for distinguishing adulteration in tea grids. This test is confirmed both the genotyping of named PCR DNA fragments and the sequencing of DNA barcoding markers for each sample [18]. Castro et al utilized a multi-faceted DNA barcoding way to deal with build up a quick and solid convention for the taxonomic classification and affirmation of herbal imbuelements in teas. The *Camellia sinensis* (L.) Kuntzethe customary tea plant, was chosen,as a model plant for this protocol. Firstly, candidate DNA barcoding markers will be selected based on the following four criteria opt for an ideal nucleotide sequences barcode were considered:

- (1) Highly proficient amplification,
- (2) High-quality sequences
- (3) An exhaustive sequence database publicly available, and
- (4) High species discrimination proficiency [15, 31, 43, 58, 83].

The accompanying two molecular strategies utilized for the DNA Verity Test (DVT): sequencing through Sanger science and genotyping of a fluoresced amplified fragment employing capillary electrophoresis. DVT is a normalized protocol of DNA barcoding description (DNA Verity Test, DVT) that was created to increment both proficiency and quickness.



Both together 32 dark and green tea samples of *Camellia sinensis* was proficiently explored, after the optimization of the DVT convention, by the author Castro et al.

E. High-resolution melting curve analysis (HRM).

To overcome certain limitations to the above-described techniques. Recently, a few literary works report another procedure in particular high resolution melting analysis (HRM) in herbal grown medicine plants identification, and these examinations uncovered a capability of HRM consolidated DNA standardized identification markers adequately recognize restorative plants, just as to identify furthest reaches of defilement in commercial tea items [37, 67]. HRM innovation shows nucleic acid samples based on their disassociation behavior, utilizing direct melting to distinguish minor sequence changes in PCR-enhanced tea samples (Sun W et al., 2016). These unmistakable highlights are recognized using DNA-explicit dyes, top of the line instrumentation, and required analysis programming. Tests are separated by their nucleotide composition, length, GC (guanine-cytosine) content, and strand complementary (Palais et al., 2005). A significant improvement of HRM is that the analysis is performed following the qTRPCR enhancement. Hence, it is especially reasonable for medium to high throughput enhancement. As of late, Song et al., 2015, utilized the ITS2 standardized identification joined with HRM innovation to distinguish five *Artemisia* species and verify their commercial items sold in China. DNA barcoding was utilized in blend with High Resolution Melting investigation (BarHRM) to confirm three restorative Acanthaceae species (*Acanthus ebracteate*, *Andrographis paniculata* and *Rhinacanthus nasutus*) regularly utilized in Thailand [66].

V. CONCLUSION:

After industrial packing, the species from which the tea products are acquired are morphologically unrecognizable. In this manner, we can't know with sureness utilizing conventional techniques if the source plants are those revealed in the recorded integrates. Thus, there is tremendous interest in the fast and effective techniques for validating and recognizing likely biological contaminants in food ventures. This audit features a portion of the molecular DNA based strategies that have demonstrated utility both in verification just as in the location of sullyng or tainting species. We accept that particular molecular markers will be fundamental for the confirmation of natural contaminants. Polymorphic marker districts, for example, the ITS area, have been appeared to recognize even firmly related species. Other loci that presently can't seem to be recognized for their utility can likewise be contaminated. Regularly such plant distinguishing proof initially includes the enhancement of the DNA locale of intrigue and afterward recognizable proof by affirming the nearness of polymorphisms explicit to the objective species. It might be important to utilize different DNA loci to check personality. There is a basic need to evaluate how much intra-species and between species variety exists in these sub-atomic markers, so exact investigations can be made. Indeed, even this modest quantity of variety could hugely affect the viability of strategies that can recognize SNPs. Recognizable proof of

the varieties inside an animal category will permit touchy advancements, for example, microarrays and species-explicit PCR to be executed and will empower DNA groupings acquired from botanicals to be contrasted with those in databases for Identification.

There are extraordinary arrangements of enthusiasm for the utilization of SNPs as a method for screening for illness in people has prompted the dynamic advancement of innovations to break down and distinguish these SNPs. Recently DNA variety test can distinguish corruption in tea lattices, here the author recommends rps7-trnV(GAC) can be utilized to segregate *C.sinensis* from *C.pubicosta* rather ITS2.

The different examination demonstrated DNA barcoding as a compelling method for testing the integrity and the genuineness of commercial tea items. Barcoding loci viz., matK, psbA-trnH, rbcL, rpoC1 were utilized to identify plant-based adulteration of commercial teas. Cloning and sequencing of the specific standardized tag loci band should be done to confirm the outcomes. Expansive scope usage of plant DNA barcoding may require calculations that home indexed lists in the setting of standard plant names and character-based keys for recognizing intently related species. Be that as it may, DNA barcoding was utilized in a joint effort with High Resolution Melting analysis (BarHRM) to clarify different species-based defilement. The rbcL standardized identification was picked for use in preliminary structures for HRM analysis to deliver standard softening profiles of the picked species. Dissolving information from the HRM measure utilizing the planned rbcL primers demonstrated that the picked species could be recognized from one another. The Bar-HRM technique grew effectively demonstrated helpful in supporting in the recognizable proof and verification of comparative species in the commercial tea sample. Later on, species validation through Bar-HRM could be utilized to advance customer trust, just as raising the quality of tea items.

REFERENCE:

1. 164th annual report Tea board, India. 2017-18. <http://www.teaboard.gov.in/TEABOARDPAGE/ODA>=
2. Ahmed, S., et al. Pu-erh tea tasting in Yunnan, China: correlation of drinkers' perceptions to phytochemistry. *J. Ethnopharmacol.* 2010. 132, 176-185.
3. Alemu Girma Tura, Dechasa Bersissa Seboka. Review on Honey Adulteration and Detection of Adulterants in Honey. *International Journal of Gastroenterology.* Vol. 4, No. 1, 2019, pp. 1-6. doi: 10.11648/j.20200401.11.
4. Andrews, E. (2008). Devouring the Gothic: Food and the Gothic body. <http://dspace.stir.ac.uk/handle/1893/375>
5. Balasaravanan, T., Pius, P. K., Kumar, R. R., Muraleedharan, N. & Shasany, A. K. Genetic diversity among south Indian tea germplasm (*Camellia sinensis*, *C. assamica*, and *C. assamica* spp. *Lasiocalyx*) using AFLP markers. *Plant Sci.* 2003. 165, 365-372.
6. Baldwin, B., Sanderson, M., Porter, J., Wojciechowski, M., Campbell, C. and Donoghue M. 1995. The ITS region of nuclear ribosomal DNA: a valuable source of evidence on angiosperm phylogeny. *Ann. Missouri Bot. Garden.* 82:247-277.
7. Barbin DF, ElMasry G, Sun DW, Allen P Predicting quality and sensory attributes of pork using near-infrared hyperspectral imaging. *Anal Chim Acta*, 2012. 719: 30-42.



8. Bibi, T., Ahmad, M., Bakhsh Tareen, R., Mohammad Tareen, N., Jabeen, R., Rehman, S.-U., Sultana, S., Zafar, M., & Yaseen, G. Ethnobotany of medicinal plants in district Mastung of Balochistan province-Pakistan. *Journal of Ethnopharmacology*, 2014. 157, 79–89. <https://doi.org/10.1016/j.jep.2014.08.042>.
9. Blattner, F.R. Direct amplification of the entire ITS region from poorly preserved plant material using recombinant PCR. *Biotechniques*, 1999. 27:1180-1186.
10. Blohm, D.H. and Guiseppi-Elie A. New developments in microarray technology. *Curr. Opin. Biotech.*, 2001. 12:41–47.
11. Borresen, A. L., E. Hovig, and A. Brogger. Detection of base mutations in genomic DNA using denaturing gradient gel electrophoresis (DGGE) followed by transfer and hybridization with gene-specific probes. *Mutat. Res.* 202:77-83.
12. Cao, H., Sasaki, Y., Fushimi, H. and Komatsu, K. 2001. Molecular analysis of medicinally-used Chinese and Japanese Curcuma based on 18S rRNA gene and trnK gene sequences. *Biol. Pharm. Bull.* 24:1389-1394.
13. Cariello, N. F., J. K. Scott, A. G. Kat, and W. G. Thilly. 1988. Resolution of a missense mutant in human genomic DNA by denaturing gradient gel electrophoresis and direct sequencing using in vitro DNA amplification: HPRTMUnich. *Am. J. Hum. Genet.* 42:726-734.
14. Carles, M., Cheung, M.K.L., Moganti, S., Dong, T.T.X., Tsim, K.W., Ip, N.Y. and Sucher, N.J. 2005. A DNA microarray for the authentication of toxic traditional Chinese medicinal plants. *Planta Med.* 71:580-584.
15. CBOL, Plant Working Group. A DNA barcode for land plants. *Proc Natl Acad Sci USA.* 2009; 106: 12794–12797.
16. Ceresa, M. (1996) Diffusion of tea-drinking habit in pre-Tang and early Tang period. *Asiatica Venetiana.* 1996. 1, 19–25.
17. Chang, H. T. A taxonomy of the genus *Camellia*. *Acta Sci. National Uni.v Sunyatseni, Monog.* 1981, Series 1, 1–180.
18. De Castro O, Comparone M, Di Maio A, Del Guacchio E, Menale B, Troisi J, et al. What is in your cup of tea? *DNA Verity Test* to characterize black and green commercial teas. *PLoS ONE*, 2017, 12(5): e0178262 <https://doi.org/10.1371/journal.pone.0178262>
19. Deng SG, Xu YF, Li L, Li XL, He Y A feature-selection algorithm based on support vector machine-multiclass for hyperspectral visible spectral analysis. *J Food Eng.* 2013, 119: 159–166.
20. Ding, X., Wang, Z., Zhou, K., Xu, L., Xu, H. and Wang, Y. 2003. Allele-specific primers for diagnostic PCR identification of *Dendrobium officinale*. *Planta Med.* 69:587-588.
21. Fan Zhu, Ratchaneekorn Sakulnak, Sunan Wang. Effect of black tea on antioxidant, textural, and sensory properties of Chinese steamed bread. *Food Chemistry* Volume 194, 1 March 2016, Pages 1217-1223.
22. Fu, R.-Z., Wang, J., Zhang, Y.-B., Wang, Z.-T., But, P. P.-H., Li, N. and Shaw, P.-C. 1999. Differentiation of medicinal *Codonopsis* species from adulterants by Polymerase Chain Reaction-Restriction Fragment Length Polymorphism. *Planta Med.* 65:648-650.
23. Galimberti A, De Mattia FA, Losa, Bruni I. DNA barcoding as a new tool for food traceability, 2013. *Food Res Int* 50:55–63.
24. Gray, M., A. Charpentier, K. Walsh, P. Wu, and W. Bender. 1991. Mapping point mutations in the *Drosophila* rosy locus using denaturing gradient gel blots. *Genetics* 127:139-149.
25. Ha, W.-Y., Shaw, P.-C., Liu, J., Yau, F. C.-F. and Wang, J. 2002. Authentication of *Panax ginseng* and *Panax quinquefolius* using amplified fragment length polymorphisms (AFLP) and directed amplification of minisatellite region DNA (DAMD). *J. Agric. Food Chem.* 50:1871-1875.
26. Hacia, J.G., Brody, L.C., Chee, M.S., Fodor, S.P. and Collins, F.S. 1996. Detection of heterozygous mutations in BRCA1 using high density oligonucleotide arrays and two-colour fluorescence analysis. *Nature Genet.* 14:441-447.
27. Hayashi K. 1991. PCR-SSCP: a simple and sensitive method for detection of mutations in the genomic DNA. *PCR Methods Appl.* 1:34-38.
28. H. B. Sowbhagy, Chemistry, Technology, and Nutraceutical Functions of Cumin (*cuminum cyminum L*): An Overview, *Critical Reviews in Food Science and Nutrition*, 2013, 53:1, 1-10, DOI: 10.1080/10408398.2010.500223.
29. Hebert, P. D. & Gretory, T. R. The promise of DNA barcoding for taxonomy, 2005. *Syst Biol.* 54, 852–859.
30. Hebert, Paul D. N.; Cywinska, Alina; Ball, Shelley L.; deWaard, Jeremy R. 2003. "Biological identifications through DNA barcodes" *Proceedings of the Royal Society of London. Series B: Biological Sciences.* 270 (1512): 313–321.
31. Hollingsworth PM, Graham SW, Little DP. Choosing and Using a Plant DNA Barcode. *PLoS ONE.* 2011; 6(5): e19254.
32. Huang M, Wang QG, Zhang M, Zhu QB Precision of color and moisture content for vegetable soybean during drying using hyperspectral imaging technology, 2014. *J Food Eng* 128: 24–30.
33. Ibrahim A. Arif, Mohammad A. Bakir, Haseeb A. Khan, Ahmad H. Al Farhan, Ali A. Al Homaidan, Ali H. Bahkali, Mohammad Al Sadoon, Mohammad Shobrak. A brief review of molecular techniques to assess plant diversity. *Int J Mol Sci.* 2010; 11(5): 2079–2096. Published online 2010 May 10. doi: 10.3390/ijms11052079.
34. Iqbal A, Sun DW, Allen P Predicting of moisture, color and pH in cooked, pre-sliced turkey hams by NIR hyperspectral imaging system. *J Food Eng.* 2013, 117: 42–51.
35. Isaac R Rukundo, Mary-Grace C Danao, Curtis L Weller, Randy L Wehling, Kent M Eskridge. Use of a handheld near infrared spectrometer and partial least squares regression to quantify metanil yellow adulteration in turmeric powder. *Journal of Near Infrared Spectroscopy*, 2020. 28:2, pages 81-92.
36. Johnson, L.A. and Soltis, D.E. 1995. Phylogenetic interference in Saxifragaceae sensu stricto and Gilia (Polemoniaceae) using matK sequences. *Ann. Missouri Bot. Gard.* 82:149-175.
37. Kalivas, A. et al. DNA barcode ITS2 coupled with high resolution melting (HRM) analysis for taxonomic identification of *Sideritis* species growing in Greece, 2014. *Mol. Biol. Rep.* 41, 5147–5155.
38. Karimi Y, Maftoonzad N, Ramaswamy HS, Prasher SO, Marcotte M. Application of hyperspectral technique for color classification avocados subjected to different treatments, 2012. *Food Bioprocess Tech* 5: 252–264.
39. Kim, D.-H., Heber, D. and Still, D.W. 2004. Genetic diversity of *Echinacea* species based on amplified fragment length polymorphisms. *Genome.* 47:102-111.
40. Lai, J.-A., Yang, W.-C. & Hsia, J.-Y. An assessment of genetic relationships in cultivated tea clones and native wild tea in Taiwan using RAPD and ISSR markers. *Bot. Bull. Acad. Sin.* 2001. 42, 93–100.
41. Lerman, L. S., S. G. Fischer, I. Hurley, K. Silverstein, and N. Lumelsky. 1984. Sequence-determined DNA separations. *Annu. Rev. Biophys. Bioeng.* 13:399-423.
42. LeRoy, A., Potter, E., Woo, H.-H., Heber, D. and Hirsch, A.M. 2002. PCR-based identification of degraded DNA and monitoring of flavonoids isolated from alfalfa and red clover commercial preparations used as botanicals in dietary supplements. *J. Agric. Food Chem.* 50:5063-5069.
43. Li X, Yang Y, Henry RJ, Rossetto M, Wang Y, Chen S. Plant DNA barcoding: from gene to genome. *Biol Rev Camb Philos Soc.* 2015; 90(1): 157–166.
44. Li, M. et al. Authentication of the anti-tumor herb *Baihuasheshicao* with bioactive marker compounds and molecular sequences, 2010. *Food Chem.* 119, 1239–1245.
45. Li, X., Patron, A., Tachdjian, C., Xu, H., Li, Q., Pronin, A., Servant, G., Zhang, L., Brady, T., Darmohusodo, V., Arellano, M., Selchau, V., Ching, B. W., Karanewsky, D. S., Brust, P., Ling, J., Zhao, W., & Priest, C. Compounds that inhibit (block) bitter taste in composition and methods of making same (United States Patent No. US20130183252A1). 2013, <https://patents.google.com/patent/US20130183252/en>
46. Li, Y.F., Li, Y.X., Lin, J., Xu, Y., Yan, F., Tang, L. and Chen, F. 2002. Identification of bulb from *Fritillaria cirrhosa* by PCR with specific primers. *Planta Med.* 69:186-188.
47. Li, H.L. The domestication of plants in China: eco geographical considerations. In: Keightley, D. N., editor. *The Origins of Chinese Civilization*. Berkeley: University of California Press, 1982. pp. 21–64.
48. Liu, Y., Pu, H., Sun, D.-W., Hyperspectral imaging technique for evaluating food quality and safety during various processes: A review of recent applications, *Trends in Food Science & Technology*, 2017. doi: 10.1016/j.tifs.2017.08.013.
49. Lum, M., Potter, E., Dang, T., Heber, D., Hardy, M. and Hirsch, A.M. 2005. Identification of botanicals and potential contaminants through RFLP and sequencing. *Planta Med.* Published online July 29, 2005.
50. Madhusudana, P. P., Uma, K. N., & Pandey, D. Heavy metals occurrence in the tissues of marine prawn *Penaeus monodon* (Fabricius 1798) and water along the coastline of Tamil Nadu (Chennai). *Asian Journal of Advances in Research*, 2020. 3(2), 23-28.



51. Magoma, G.N., Wachira, F.N., Obanda, M., Imbuga, M. & Agong, S.G. The use of catechins as biochemical markers in diversity studies of tea (*Camellia sinensis*). *Genet. Resources Crops. Evol.* 2000. 47, 107–114.
52. Manning and Soon. 2014 Developing systems to control food adulteration. *Food policy* 49 (2014) 23–32.
53. Michael Gundry and Jan Vijg 2012. “Direct mutation analysis by high-throughput sequencing: From germline to low-abundant, somatic variants”. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis Volume 729, Issues 1–2, 3 January 2012, Pages 1–15.*
54. Micozzi, M. S. *Fundamentals of Complementary, Alternative, and Integrative Medicine—E-Book.* 2018, Elsevier Health Sciences.
55. Mihalov, J.J., Marderosian, A.D. and Pierce, J.C. 2000. DNA identification of commercial ginseng samples. *J. Agric. Food Chem.* 48:3744–52.
56. Miller, S. E. DNA barcoding and the renaissance of taxonomy. *P. Natl. Acad. Sci. USA* 104, 4775–4776.
57. Ming, T. & Zhang, W. The evolution and distribution of genus *Camellia*. *Acta Botanica Yunnanica.* 1996. 18, 1–13.
58. Mishra P, Kumar A, Nagireddy A, Mani DN, Shukla AK, Tiwari R, Sundaresan V. DNA barcoding: an efficient tool to overcome authentication challenges in the herbal market. *Plant Biotechnol J.* 2016; 14 (1): 8–21. <https://doi.org/10.1111/pbi.12419> PMID: 26079154.
59. Muhammad Azhar Nadeem, Muhammad Amjad Nawaz, Muhammad Qasim Shahid, Yıldız Doğan, Gonul Comertpay, Mehtap Yıldız, Rüşti Hatipoğlu, Fiaz Ahmad, Ahmad Alsaleh, Nitin Labhane, Hakan Özkan, Gyuhwa Chung & Faheem Shehzad Baloch DNA molecular markers in plant breeding: current status and recent advancements in genomic selection and genome editing, *Biotechnology & Biotechnological Equipment*, 2018, 32:2, 261–285, DOI: 10.1080/13102818.2017.1400401.
60. Mukherjee, P. K., Pitchairajan, V., Murugan, V., Sivasankaran, P. & Khan, Y. Strategies for revitalization of traditional medicine. 2010, *Chin. Herb. Med.* 2, 1–15.
61. Myers, R. M., S. G. Fischer, L. S. Lerman, and T. Maniatis. 1985. Nearly all single base substitutions in DNA fragments joined to a GC-clamp can be detected by denaturing gradient gel electrophoresis. *Nucleic Acids Res.* 13:3131–3145.
62. Nelson, L. A., Wallman, J. F., & Dowton, M. Identification of forensically important *Chrysomya* (Diptera: Calliphoridae) species using the second ribosomal internal transcribed spacer (ITS2). *Forensic Science International*, 2008, 177(2), 238–247. <https://doi.org/10.1016/j.forsciint.2008.01.009>
63. Newmaster SG, Grguic M, Shanmughanadhan D, Ramalingam S. DNA barcoding detects contamination and substitution in North American herbal products. *BMC Med* 11:222–35.
64. Newton, C.R., Graham, A., Heptinstall, L.E., Powell, S.J., Summers, C., Kalsheker, N., Smith, J.C. and Markham, A.F. 1989. Analysis of any point mutation in DNA. The amplification refractory mutation system (ARMS). 2013, *Nuc. Acids Res.* 17:2503–2516.
65. Orita M, Iwahana H, Kanazawa H, Hayashi K and Sekiya T. 1989. Detection of polymorphisms of human DNA by gel electrophoresis as single-strand conformation polymorphisms. *Proc. Natl. Acad. Sci. USA.* 86: 2766–2770.
66. Osathanunkul M, Madesis P, de Boer H Bar-HRM for Authentication of Plant-Based Medicines: Evaluation of Three Medicinal Products Derived from Acanthaceae Species. 2015, *PLoS ONE* 10 (5).
67. Osathanunkul, M., Suwannapoom, C., Osathanunkul, K., Madesis, P. & de Boer, H. Evaluation of DNA barcoding coupled high resolution melting for discrimination of closely related species in phytopharmaceuticals. 2016, *Phytomedicine* 23, 156–165.
68. Palais, R., Liew, M. & Wittwer, C. Quantitative heteroduplex analysis for single nucleotide polymorphism genotyping. *Anal. Biochem.* 346, 167–175.
69. Ramsar, J., Lopez-Peralta, C., Wetzler, R., Weising, K. and Kahl, G. 1996. Genomic variation and relationships in aerial yam (*Dioscorea bulbifera* L.) detected by random amplified polymorphic DNA. *Genome.* 39:17–25.
70. Ryan, G. T. DNA barcoding does not compete with taxonomy. 2005, *Nature* 434, 1067–1067.
71. S K V Bharathi, A Sukitha, J A Moses & C Anandharamkrishnan. Instrument-based detection methods for adulteration in spice and spice products – 2018, A review. *Journal of Spices and Aromatic Crops Vol. 27 (2) : 106–118.*
72. Saiki, R. K., S. J. Scharf, F. Faloona, K. B. Mullis, G. T. Horn, H. A. Erlich, and N. Arnheim. 1985. Enzymatic amplification of beta-globin genomic sequences and restriction site analysis for diagnosis of sickle cell anemia. *Science* 230:1350–1354.
73. Saleena Mathew Maya Raman Manjusha Kalarikkathara Parameswaran Dhanya Pulikkottil Rajan Techniques used in fish and fisheries products analysis *Fish and Fishery Products Analysis* pp 263–360.
74. Sangita Bansal, Apoorva Singh, Manisha Mangal, Anupam K. Mangal & Sanjiv Kumar Food adulteration: Sources, health risks, and detection methods, *Critical Reviews in Food Science and Nutrition*, 2017, 57:6, 1174–1189.
75. Sasaki, Y., Fushimi, H., Cao, H., Cai, S.-Q., and Komatsu, K. 2002. Sequence analysis of Chinese and Japanese Curcuma drugs on the 18S rRNA gene and trnK gene and the application of amplification refractory mutation system for their authentication. *Biol. Pharm. Bull.* 25:1593–1599.
76. Schwieger, F. and Tebbe, C.C. 1998. A new approach to utilize PCR-single-strand conformation polymorphism for 16S rRNA gene-based microbial community analysis. *Appl. Environ. Microbiol.* 64: 4870–4876.
77. Semenov, G. A., Safran, R. J., Smith, C. C. R., Turbek, S. P., Mullen, S. P., & Flaxman, S. M. . *Unifying Theoretical and Empirical Perspectives on Genomic Differentiation. Trends in Ecology & Evolution.* 2019, doi:10.1016/j.tree.2019.07.008.
78. Shaw, P.-C., Ngan, F.N., But, P.P.-H., and Wang, J. 2002. Molecular markers in Chinese medicinal materials. In: Shaw, P.-C., Wang, J. and But, P.P.-H. (eds.) *Authentication of Chinese Medicinal Materials by DNA Technology.* World Scientific, Singapore. Pp. 1–23.
79. Sheffield, V. C., D. R. Cox, L. S. Lerman, and R. M. Myers. 1989. Attachment of a 40-base pair G+C-rich sequence (GC-clamp) to genomic DNA fragments by the polymerase chain reaction results in improved detection of single-base changes. *Proc. Natl. Acad. Sci. USA* 86:232–236.
80. Sheffield, V.C., Beck, J.S., Kwitek, A.E., Sandstrom, D.W., and Stone, E.M. 1993. The sensitivity of single-strand conformation polymorphism analysis for the detection of single base substitutions. *Genomics.* 16:325–32.
81. Song, M. et al. Applying high-resolution melting (HRM) technology to identify five commonly used *Artemisia* species. 2016, *Sci. Rep.* 6, 34133; doi: 10.1038/srep34133.
82. Still, D.W., Kim, D.-H. and Aoyama, N. 2005. Genetic variation in *Echinacea angustifolia* along a climatic gradient. *Ann. Bot.* <http://aob.oxfordjournals.org>.
83. Stoeckle MY, Gamble CC, Kirpekar R, Young G, Ahmed S, Little DP. Commercial teas highlight plant DNA barcode identification successes and obstacles. *Sci Rep* 2011; 1: 42.
84. Sun W, Li J-j, Xiong C, Zhao B and Chen S-j The Potential Power of Bar-HRM Technology in Herbal Medicine Identification. 2016, *Front. Plant Sci.* 7:367. doi:10.3389/fpls.2016.00367.
85. Sun, Y., Fung, K.-P., Leung, P.-C., Shi, D. and Shaw, P.-C. 2004. Characterization of medicinal *Epimedium* species by 5S rRNA gene spacer sequencing. *Planta Med.* 70:287–288.
86. Sunnucks, P., Wilson, A.C.C., Beheregaray, L.B., Zenger, K., French, J. and Taylor, A.C. 2000. SSCP is not so difficult: the application and utility of single-stranded conformation polymorphism in evolutionary biology and molecular ecology. *Mol. Ecol.* 9:1699–1710.
87. T. R. Dias, et al. White Tea (*Camellia sinensis* (L.)): Antioxidant Properties and Beneficial Health Effects. 2013, *Int J Food Sci Nutr Diet.* 2(2), 19–26. doi: <http://dx.doi.org/10.19070/2326-3350-130005>.
88. Techen, N., Crockett, S.I., Khan, I.A. and Scheffler, B.E. 2004. Authentication of medicinal plants using molecular biology techniques to compliment conventional methods. *Curr. Med. Chem.* 11:1391–1401.
89. Tsoi, P.Y., Woo, H.S., Wong, M.S., Chen, S.L., Fong, W.F., Xiao, P.G. and Yang, M.S. 2003. Genotyping and species identification of *Fritillaria* by DNA chips. *Yao Xue Xue Bao.* 38:185–190.
90. Vos, P., Hogers, R., Bleeker, M., Reijnders, M., van de Lee, T., Hornes, M., Frijters, A., Pot, J., Peleman, J., Kuiper, M. and Zabeau, M. 1995. AFLP: a new technique for DNA fingerprinting. *Nuc. Acids Res.* 23:4407–4414.
91. Wachira, F.N., Waugh, R., Hackett, C.A. and Powell, W. 1995. Detection of genetic diversity in tea (*Camellia sinensis*) using RAPD markers. *Genome.* 38:201–210.



92. Wang, C.-Z., Li, P., Ding, J.Y., Jin, G.-Q. and Yuan, C.-S. 2005. Identification of *Fritillaria pallidiflora* using diagnostic PCR and PCR-RFLP based on nuclear ribosomal DNA internal transcribed spacer regions. *Planta Med.* 71:384-386.
93. Wei X, Liu F, Qiu ZJ, Shao YN, He Y Ripeness classification of astringent persimmon using hyperspectral imaging technique. 2014, *Food Bioprocess Tech* 7: 1371–1380.
94. Weising, K., Nyborn, H., Wolff, K. and Kahl, G. 2005. DNA Fingerprinting in Plants. Principles, Methods, and Applications. CRC Press, Taylor and Francis Group, Boca Raton, FL. 444 p.
95. Wu D, Sun DW Potential of time series-hyperspectral imaging (TS-HSI) for non-invasive determination of microbial spoilage of salmon flesh. 2013, *Talanta* 111: 39–46.
96. Wu D, Sun DW, He Y Application of long-wave near infrared hyperspectral imaging for measurement of color distribution in salmon fillet. 2012, *Innov Food Sci Emerg* 16: 361–372.
97. Wu D, Wang SJ, Wang NF, Nie PC, He Y, et al. Application of time series hyperspectral imaging (TS-HSI) for determining water distribution within beef and spectral kinetics analysis during dehydration. 2013, *Food Bioprocess Tech* 6: 2943–2958.
98. Wu JH, Peng YK, Li YY, Chen JJ, Dhakal S Prediction of beer quality attributes using VIS/NIR hyperspectral scattering imaging technique. 2012, *J Food Eng* 109: 267–273.
99. Xie C, Li X, Shao Y, He Y (2014) Color Measurement of Tea Leaves at Different Drying Periods Using Hyperspectral Imaging Technique. *PLoS ONE* 9(12): 2014. e113422.
100. Xin, T. Y. et al. Survey of commercial *Rhodiola* products revealed species diversity and potential safety issues. 2015, *Sci. Rep.* 5, 8337.
101. Yang, D.-Y., Fushimi, H., Cai, S.-Q. and Komatsu, K. 2004. Polymerase chain reaction/restriction fragment length polymorphism (PCR-RFLP) and amplification refractory mutation system (ARMS) analyses of medicinally used *Rheum* species and their application for identification of *rhei* rhizome. *Biol. Pharm. Bull.* 27:661-669.
102. Yuan, M. and Hong, Y. 2003. Heterogeneity of Chinese medicinal herbs in Singapore assessed by fluorescence AFLP analysis. *Amer. J. Chin. Med.* 31:773-779.
103. Zhang, Y.-B., Wang, J., Wang, Z.-T., But, P.P.-H. and Shaw, P.-C. 2003. DNA microarray for identification of the herb of *Dendrobium* species from Chinese medicinal formulations. *Planta Med.* 69:1172-1174.
104. Zhu, S., Fushimi, H., Cai, S., and Komatsu, K. 2004. Species identification from ginseng drugs by multiplex amplification refractory mutation systems (MARMS). *Planta Med.* 70:189-192.



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